

# Detection of Brazilian Hantavirus by Reverse Transcription Polymerase Chain Reaction Amplification of N Gene in Patients with Hantavirus Cardiopulmonary Syndrome

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*We report a nested reverse transcription-polymerase chain reaction (RT-PCR) assay for hantavirus using primers selected to match high homology regions of hantavirus genomes detected from the whole blood of hantavirus cardiopulmonary syndrome (HCPS) patients from Brazil, also including the N gene nucleotide sequence of Araraquara virus. Hantavirus genomes were detected in eight out of nine blood samples from the HCPS patients by RT-PCR (88.9% positivity) and in all 9 blood samples (100% positivity) by nested-PCR. The eight amplicons obtained by RT-PCR (P1, P3-P9), including one obtained by nested-PCR (P-2) and not obtained by RT-PCR, were sequenced and showed high homology (94.8% to 99.1%) with the N gene of Araraquara hantavirus. Although the serologic method ELISA is the most appropriate test for HCPS diagnosis, the use of nested RT-PCR for hantavirus in Brazil would contribute to the diagnosis of acute hantavirus disease detecting viral genomes in patient specimens as well as initial genomic characterization of circulating hantaviruses.*

Key words: hantavirus cardiopulmonary syndrome - nested RT-PCR - N gene of hantavirus

Since it was described in 1993, hantavirus cardiopulmonary syndrome (HCPS) has been reported in virtually the entire American Continent. Presently, most of the reported HCPS cases are from South America, where the Andes strain of hantavirus is responsible for the majority of the cases (Nichol et al. 1993, Padula et al. 2000a).

*Hantavirus* is a genus within the *Bunyaviridae* family. These viruses are enveloped negative-stranded RNA virus with a tripartite negative-sense RNA genome whose segments are designated L, M, S, respectively coding for the polymerase associated to the virion, two surface glycoproteins (G1 and G2), and the nucleocapsid protein (N) (Schmaljohn 1996).

The evolution of the American hantaviruses has an intimate relationship with the evolution of rodent reservoirs, which are classified in *Rodentia* order, *Muridae* family and *Sigmodontinae* sub-family (Schmaljohn & Hjelle 1997, Zhao & Hay 1997). Humans are infected through the respiratory route by contact with rodent excretions and secretions. However, during an outbreak in Argentina caused by Andes virus, person-to-person transmission was described (Wells et al. 1997).

A hantavirus was isolated for the first time in Brazil from *Rattus norvegicus* in the northern region of the state of Pará, in 1982 (Le Duc et al. 1985). In 1993, three HCPS cases were detected in members of one family who lived in a farm in close contact with wild rodents, in Jucituba, SP, Southeast Brazil. Two of these three patients died.

Between 1993 and December 2003, about 329 HCPS cases were reported in Brazil (Iversson 1996, Funasa 2003). HCPS occurred in people of both sexes, aged 9 months to 66 years, with a 40% case fatality rate. Most of the HCPS cases in Brazil occurred in the Southern and Southeastern regions (Funasa 2003).

Several hantaviruses have been detected in South America, such as Andes (Lopez et al. 1997), Laguna Negra (Johnson et al. 1997), Rio Mamoré (Bharadwaj et al. 1997), and Caño Delgadito (Fulhorst et al. 1997). To assess the degree of genomic variability of these viruses, partial nucleotide sequences of the M and S viral RNA segments were analyzed (Padula et al. 2000a). These studies showed that HCPS patients from Argentina, Chile, Paraguay, Uruguay were infected with the lineages Andes or Laguna Negra. Rio Mamore, a hantavirus isolated in Iquitos, Peru, showed a closer relationship with Laguna Negra than with Andes virus based on phylogenetic analysis of the complete S segment (Bharadwaj et al. 1997).

Partial nucleotide sequences of the N, G1, G2 genes obtained from acute-phase serum samples from 3 HCPS patients from Castelo dos Sonhos, state of Mato Grosso, in Central Brazil, and from Araraquara and Franca, state of São Paulo, in Southeast Brazil, were analyzed (Johnson et al. 1999). In the phylogenetic analysis based on nucleotide and deduced amino acid sequences, the identity between Araraquara and Franca viruses revealed to be high and the three Brazilian hantaviruses appeared to be more closely related to the Argentinean Andes virus.

HCPS has become a public health problem in Ribeirão Preto City and neighboring towns in the state of São Paulo, Brazil (Johnson et al. 1999, Figueiredo et al. 1999). In the last 5 years, 29 cases of HCPS were reported in the region of Ribeirão Preto with a 48% fatality rate (Funasa 2003). Clinical samples from nine of these patients were analyzed in the present study.

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The diagnosis of hantavirus infections in Brazil has been confirmed only by the detection of IgM antibodies against Sin Nombre virus by ELISA serologic test using recombinant N protein provided by Center for Diseases Control (CDC, US), or immunohistochemistry carried out at Instituto Adolfo Lutz, São Paulo (Ksiasek et al. 1995, Zaki et al. 1995). Also was used in the ELISA a recombinant N protein from Andes virus provided by Carlos Malbran Institute, Buenos Aires, Argentina (Padula et al. 2000b). These have been the only available tests, and it is done in only three centers (Adolfo Lutz, Evandro Chagas, Oswaldo Cruz Institutes), due to difficulties to obtain hantavirus antigens that are not produced in the country. As an alternative diagnostic method, virus detection in patient specimens can be performed by reverse transcription-polymerase chain reaction (RT-PCR) amplification, an efficient method first described for HCPS diagnosis in 1993 (Le Guenno 1993, Kim et al. 1994, Johnson et al. 1999, Garin et al. 2001). Although this molecular technique has been applied as an alternative diagnostic method, serologic test (ELISA) can and must be the method of choice for the diagnosis of this zoonosis, because in the public health, ELISA is the most appropriate test for HCPS diagnosis. We report here the detection of Brazilian hantavirus genome in human blood samples from HCPS patients by a nested RT-PCR. In addition, we report a phylogenetic analysis of nucleotide sequences of hantavirus N gene obtained from HCPS patients.

#### MATERIALS AND METHODS

*Blood samples of HCPS patients and controls for the nested RT-PCR* - Whole blood samples from nine patients (P) with HCPS from Ribeirão Preto area, state of São Paulo, Brazil (Table I), were processed for hantavirus genome detection by nested RT-PCR. All 9 P had HCPS diagnosis

confirmed by the detection of IgM antibodies to Sin Nombre virus antigens (CDC, US) in an ELISA done by the Adolfo Lutz Institute, São Paulo City, Brazil (Ksiasek et al. 1995). The blood samples remained stored for 3 months to 3 years, at  $-70^{\circ}\text{C}$ . A plasmid (pET23b) containing the N gene of Andes hantavirus was kindly supplied by Dr Brian Hjelle, New Mexico, US. This plasmid was propagated in *Escherichia coli*, purified with the Plasmid Mini Kit (Qiagen, Germany) and used as a positive control in the nested RT-PCR assay. Water and a blood sample from a patient with dengue were used as negative controls.

*RNA extraction* - RNA was extracted from 300  $\mu\text{l}$  of each hemolysed whole blood sample, which was mixed with 1 ml of Trizol LS reagent (Gibco, US), and 200  $\mu\text{l}$  of chloroform-isoamyl alcohol (24:1) according to modifications described previously by Bowen et al. (1997). Pellets were suspended in 10 to 20  $\mu\text{l}$  of diethyl pyrocarbonate (DEPC) treated water.

*RT-PCR primers* - Nucleotide sequences of the N gene of the South American hantaviruses, Andes, Rio Mamoré, Araraquara, and Castelo dos Sonhos were aligned by the Higgins and Sharp algorithm, using default stringency parameters as recommended by the DNASIS for Windows version 2.10 software (Hitachi, Japan). After alignment of the hantaviruses nucleotide sequences two primers for RT-PCR were designed in highly conserved regions in the N gene (SAHN-C and SAHN-S, Table II) to amplify a region of 264 nucleotides.

*RT-PCR* - For the reverse transcription (RT), a 20  $\mu\text{l}$  volume reaction mixture containing 5  $\mu\text{l}$  of the RNA extract, 1  $\mu\text{l}$  of 2.5 mM deoxyribonucleoside triphosphates (dNTPs), 0.3  $\mu\text{M}$  of the SAHN-C primer, and 4  $\mu\text{l}$  of RT 5x buffer (10mM Tris pH 8.9, 1.5 mM  $\text{MgCl}_2$ , 80 mM KCl) was prepared. The mixture was overlaid with two drops of min-

TABLE I

Hantavirus cardiopulmonary syndrome patients whose blood samples were tested by nested-polymerase chain reaction

Patient	Sex	Age (years)	Risk activity	Disease onset	Sampling day	Evolution
1	M	55	Agricultural worker	May, 1998	3 days <sup>a</sup>	Death
2	M	38	Farmer	June, 1998	3 days	Death
3	M	29	Truck driver	May, 1999	5 days	Survived
4	M	26	Farmer	May, 1999	3 days	Death
5	F <sup>c</sup>	13	Student	June, 1999	8 days	Survived
6	M	21	Agricultural worker	June, 1999	9 days	Survived
7	M	39	Agronomist	April, 2001	15 days	Survived
8	F	31	Nurse	August, 2002	3 days	Death
9	M	41	Bootmaker	April, 2002	4 days	Survived

M: male, F: female; <sup>a</sup>: sample collection days after the onset of symptoms

TABLE II

Primers used in the Brazilian hantavirus reverse transcription polymerase chain reaction (PCR) and nested-PCR

Primer	Nucleotide sequence	Annealing site
SAHN-S (sense)	GATGAATCATCCTTGAACCTTAT	Nucleotide 477 to 454 <sup>a</sup>
SAHN-C (complementary)	CAAAACCAGTTGATCCAACAGGG	Nucleotide 213 to 236 <sup>a</sup>
SAHNinternal-S (sense)	GTTGACAGCATATATAAGGCCTT	Nucleotide 391 to 414
SAHNinternal-C (complementary)	GATATGGGAATGTGCTGGATGT	Nucleotide 277 to 298

<sup>a</sup>: annealing site based on Araraquara virus.

eral oil and heated at 95°C for 3 min, in order to produce RNA linearization. After temperature reduction to 4°C, 10 U of RNase inhibitor (Pharmacia, US) and 10 U of reverse transcriptase MMLV (Pharmacia) were added to the RT reaction mixture, which was incubated for 2 h at 37°C. The resulting cDNA was used as a template in the subsequent PCR reaction. For the PCR, a 50 µl mixture containing 0.3 µM of each SAHN-C and SAHN-S primers, 1 µl of 2.5 mM of the dNTPs, 3 µl of the RT product, and 5 µl of the 10x buffer (10 mM Tris pH 8.9, 1.5 mM MgCl<sub>2</sub>, 80 mM KCl) was prepared. Finally, after heating at 80°C, 1 U of thermostable Taq DNA polymerase (Pharmacia) was added and the mixture was submitted to 35 cycles in a thermal cycler (Iwaki, Japan), at the following temperatures: 95°C for 30 s, 55°C for 45 s, and 72°C for 180 s. Finally, the mixture was held for 5 min at 72°C. Eight µl of the amplified DNA was electrophoresed in 1.7% agarose gels in TAE buffer and stained with ethidium bromide (1 µg/ml in water solution). DNA was visualized by using a 312 nm UV transilluminator and camera. Amplicon sizes were determined by comparison with a 100 base pair (bp) DNA ladder (Pharmacia).

**Analysis of nucleotide sequences and primers selection for the nested-PCR** - Amplicon nucleotide sequences from the HCPS P and GenBank sequences of S segment from Hantaan virus (U37768), Andes virus (AF325966), Sin Nombre virus (L37904), Laguna Negra virus (AF005727), Rio Mamoré virus (U52136), Araraquara virus (AF307325), and Castelo dos Sonhos virus (AF307324) were aligned by the Higgins and Sharp algorithm, using default stringency parameters as recommended by the DNASIS for Windows version 2.10 software (Hitachi, Japan). Nested-PCR internal primers (SAHNinternal-S and SAHNinternal-C) shown in Table II were chosen from the aligned nucleotide sequences in high homology regions and matching between nucleotides 277 to 298 and 391 to 414 to produce a 140 bp amplicon. The phylogenetic analysis of the aligned nucleotide sequences of patient samples was performed using the same software, by applying the UPGMA method (Unweighted Pair Group Method with Arithmetic Mean).

**Nested-PCR** - Nested-PCR was carried out starting with 2 µl of the amplicons obtained in the first round of RT-PCR in a reaction mixture including 1 U of thermostable Taq DNA polymerase (Pharmacia), 5 µl of 10 x buffer, 0.3 µM of each SAHNinternal-C and SAHNinternal-S primer, and DEPC treated water completing a 50 µl volume. Twenty-five cycles of amplification were performed with the same parameters as for the first round of PCR.

**Nucleotide sequencing of RT-PCR and nested PCR amplicons** - The 264 bp amplicons from RT-PCR were subjected to 25 re-amplification cycles of PCR at 95°C for 30 s, 55°C for 45 s, and 72°C for 180 s. The amplicons were purified using Wizard PCR preps (Promega, US) and 7.5 µl of the purified DNAs were sequenced with the Thermo Sequenase CY5.5 terminator kit (Amersham-Pharmacia, England), using 1 µM of each SAHN-C and SAHN-S primers and fluorescein labeled nucleotides. The products were run in a denaturing polyacrylamide sequencing gel in a personal seq 4 x 4 sequencer (Pharmacia), for 35 min.

One 140 bp amplicon from P-2, obtained by nested-PCR, was sequenced as described above. This amplicon was selected because it was not possible to obtain the 264 bp amplicon of P-2 in the RT-PCR.

## RESULTS

**RT-PCR** - Amplicons of approximately 264 base pairs (bp) were obtained at the first round of PCR from eight out of nine blood samples from HCPS patients (88.9% positivity, Fig. 1A). Amplicons of approximately 140 bp were obtained by nested-PCR from all blood samples (Fig. 1B). The negative control blood sample of the patient infected with dengue was tested by RT-PCR and by nested PCR without showing amplicon.

**Nucleotide sequences** - The alignment of nucleotide sequences of the eight amplicons obtained by RT-PCR and one amplicon obtained by nested-PCR, showed matching percentages from 94.8 to 100% among themselves. The nine amplicons also showed 82.6% matching with Andes virus, 78.4% with Rio Mamoré virus and Laguna Negra virus, and 98.6% with Araraquara virus, as shown in the dendrogram at Fig. 2. These nine amplicons from HCPS patient blood samples are highly homologous and probably belong to Araraquara virus or a very close related virus, as shown in Fig. 3.

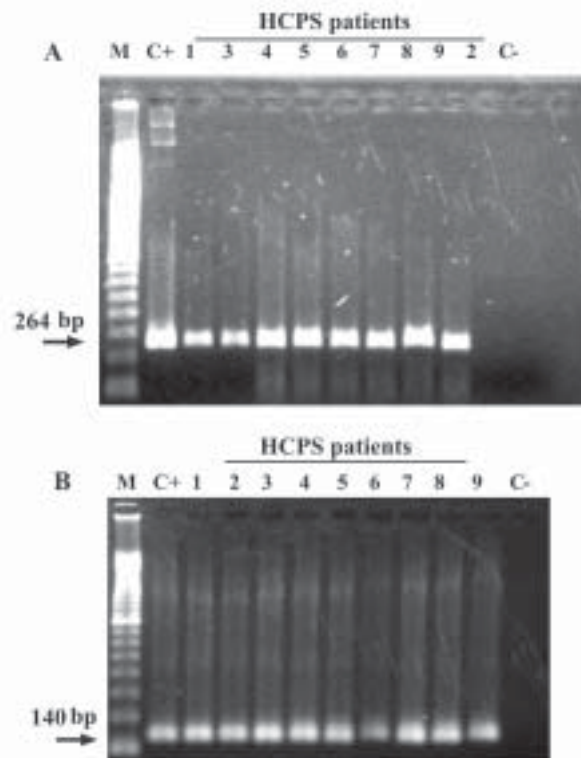


Fig. 1A: agarose gel showing amplicons having 264 bp obtained in the reverse transcription polymerase chain reaction from the sera of eight of nine hantavirus cardiopulmonary syndrome (HCPS) patients (P1; P3-P9); the amplicon from patient 2 (P-2) was not detectable. In both gels (A and B), a plasmid containing the N gene of Andes virus was used as positive control (C+). The serum from one patient with dengue was used as negative control (C-); B: agarose gel showing amplicons having 140 bp obtained in the nested-PCR from the sera of all nine HCPS patients.

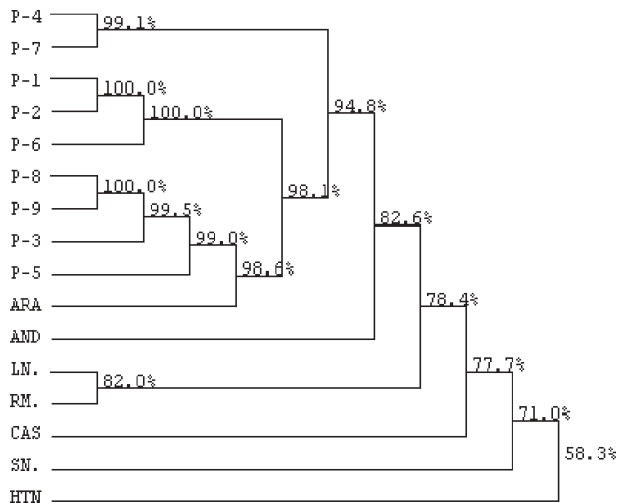


Fig. 2: phylogenetic dendrogram of nucleotide sequences of eight amplicons obtained by reverse transcription polymerase chain reaction (P1; P3-P9) and one amplicon obtained by nested-PCR (P-2) from nine hantavirus cardiopulmonary syndrome patients, showing Araraquara virus (ARA) in the same branch. Laguna Negra (LN), Rio Mamore (RM), Andes (AND) also South American hantaviruses clustered in a different branch. Castelo dos Sonhos (CAS), Sin Nombre (SN) and Hantaan (HTN) viruses were more distantly localized in the phylogenetic dendrogram. Calculated matching percentages are indicated at each branching point of the dendrogram.

## DISCUSSION

In this study, the RNA extracts from which hantavirus genome was amplified by nested-PCR came from stored whole hemolyzed blood samples collected from HCPS patients between 3 and 15 days of disease. It is remarkable that some of the samples were frozen and thawed two or three times. This suggests that this kind of sample is suitable for hantavirus genome detection by RT-PCR and that the Tryzol-LS (Gibco, US) method modified by Bowen et al. (1997) is adequate for RNA extraction from this kind of clinical sample.

The efficacy of SAHN-C and SAHN-S primers selected on the basis of sequence similarity in the N gene among Andes, Rio Mamoré, Araraquara, and Castelo dos Sonhos, all South American hantaviruses was tested for the amplification of Brazilian hantavirus genome in HCPS patients. The RT-PCR with SAHN-C and SAHN-S primers was able to amplify the hantavirus genomes in 88.9% of the tested blood samples of HCPS patients. Amplicon was not obtained in the RT-PCR and the nested-PCR from a blood sample of one patient with dengue and one water sample. The specificity of the amplicons was confirmed by the high matching percentages of their nucleotide sequences with that of Araraquara virus (94.8% to 99.1%).

In order to further enhance sensitivity a round of nested-PCR was added using primers SAHNinternalC and SAHNinternalS, selected of highly conserved regions present in hantaviral S segments internally to the RT-PCR amplicons from HCPS patients and Araraquara virus. Based on the results obtained, despite of the restricted number of HCPS tested blood samples this nested RT-PCR assay seems to be accurate and suitable for clinical use, at least in the northeastern region of the state of São Paulo, Brazil.

The nucleotide sequences of the N genes of hantavirus amplicons obtained by RT-PCR showed high homology with those of several South American hantaviruses. The highest matchings (94.8% to 99.1%) were observed with the Araraquara virus that is shown in Fig. 2 surrounded by the HCPS patients samples. This result was expected because the genome of Araraquara virus was obtained in 1996 from a HCPS patient living in Araraquara City, approximately 90 km of Ribeirão Preto (Johnson et al. 1999). Interestingly, HCPS P-1 and P-2 showed highest matching between themselves. This fact could be explained because these patients lived in the same place. Thus, we believe this virus is the causative agent of HCPS in the region of Ribeirão Preto. The other South American hantaviruses, were detected in places over 1600 km from Ribeirão Preto and clustered in a separate branch of the phylogenetic tree (Fig. 2), showing 82.6% (Andes) and 78.4% (Laguna Negra and Rio Mamoré) of homology with the hantaviruses from Ribeirão Preto associated to HCPS.

The nested-PCR with SAHN internal primers was found to be suitable for diagnosis because it detected the hantavirus genome in all the HCPS blood samples. Besides, false-positive results associated to sample contamination were not observed. Amplicons were not obtained from water or from the blood sample of the patient with dengue, as shown in Fig. 1B.

In short, this nested RT-PCR could be used as a diagnostic test of hantavirus infection. Additionally, the RT-PCR method could be used for the amplification of the entire N gene of the hantavirus contained in the blood sample of HCPS cases. This gene could be cloned and expressed in order to produce a recombinant N protein from the entire gene obtained from blood sample patients, that could be used for one more specific-ELISA. This approach could be a suitable alternative for enhancing sensitivity and specificity of the serologic test diagnosis in Brazil, using local antigens. In Brazil, ELISA is performed using American and Argentinean recombinant antigens.

Hantavirus in Brazil is becoming a public health problem since there have been more than three hundred HCPS reported cases, and the number is constantly increasing. Besides, other clinical presentations of hantavirus infections have probably occurred in the Northeast and South regions of Brazil (Hindrichsen et al. 1993, Flores SCC, state of Santa Catarina Health Ministry, pers. commun. 2001). For addition, high levels of antibodies to hantavirus in health populations have been reported in Brazil (Holmes et al. 2000). The use of a nested RT-PCR method approach for hantavirus in Brazil would provide an early rapid diagnosis of acute hantavirus disease as well as alternative for initial characterization of genomes of hantaviruses based on the nucleotide sequence of the amplicons obtained in this nested RT-PCR. Additionally, this nested RT-PCR method may provide an important and useful tool to improve public health system preparedness.

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		10	20	30	40	50	
ARA	1	TTGAGCCTGA	TGACCATCTA	AAGGAGAAAT	CATCACTGAG	ATATGGGAAT	50
P-1.	1	.....	.....	.....	.....	.....	50
P-2.	1	-----	-----	-----	-----	-----	50
P-3.	1	.....	.....	.....	.....	.....	50
P-4.	1	.....	.....	.....	.....	.....	50
P-5.	1	.....	.....	.....A.....	.....G.....	.....	50
P-6.	1	.....	.....	.....	.....	.....	50
P-7.	1	.....	.....	.....	.....	.....	50
P-8.	1	.....	.....	.....A.....	.....	.....	50
P-9.	1	.....	.....	.....A.....	.....	.....	50
		60	70	80	90	100	
ARA	51	GTGCTGGATG	TTAACCTAT	CGATCTTGAA	GAGCCAAGTG	GCCAGACAGC	100
P-1.	51	.....	.....C.....	.....	.....	.....	100
P-2.	51	.....	.....C.....	.....	.....	.....	100
P-3.	51	.....	.....C.....	.....	.....	.....	100
P-4.	51	.....	.....T.....	.....	.....C.....	.....G.CA.....	100
P-5.	51	.....	.....C.....	.....	.....	.....	100
P-6.	51	.....	.....C.....	.....	.....	.....	100
P-7.	51	.....A.A.....	.....T.....	.....	.....C.....	.....G.CA.....	100
P-8.	51	.....	.....C.....	.....	.....	.....	100
P-9.	51	.....	.....C.....	.....	.....	.....	100
		110	120	130	140	150	
ARA	101	AGACTGGAAG	GCAATTGGGG	CTTACATCTT	GGGTTTTCGA	ATCCCTATTA	150
P-1.	101	.....	.....A.....	.....	.....	.....C.....	150
P-2.	101	.....	.....A.....	.....	.....	.....C.....	150
P-3.	101	.....	.....	.....	.....	.....	150
P-4.	101	.....	.....A.....	.....	.....	.....	150
P-5.	101	.....	.....	.....	.....	.....	150
P-6.	101	.....	.....A.....	.....	.....	.....C.....	150
P-7.	101	.....	.....A.....	.....	.....	.....	150
P-8.	101	.....	.....	.....	.....	.....	150
P-9.	101	.....	.....	.....	.....	.....	150
		160	170	180	190	200	
ARA	151	TACTAAAGGC	CTTATATATG	CTGTCAACTC	GTGGAAGGCA	GACAGTGAAG	200
P-1.	151	.....	.....	.....	.....	.....	200
P-2.	151	.....	.....	.....	.....	.....	200
P-3.	151	.....	.....	.....C.....	.....	.....	200
P-4.	151	.....	.....	.....	.....	.....T.....	200
P-5.	151	.....	.....	.....C.....	.....	.....A.....	200
P-6.	151	.....	.....	.....	.....	.....	200
P-7.	151	.....	.....	.....	.....	.....T.....	200
P-8.	151	.....	.....	.....C.....	.....	.....	200
P-9.	151	.....	.....	.....C.....	.....	.....	200
		210	220	230	240	250	
ARA	201	GACAACAAGG	GCACTAGGAT	AAGGTTCAAG	GATGATTCA.	.....	250
P-1.	201	.....	.....	.....	.....	.....	250
P-2.	201	-----	-----	-----	-----	.....	250
P-3.	201	.....	.....	.....	.....	.....	250
P-4.	201	.....	.....	.....	.....	.....	250
P-5.	201	.....	.....	.....	.....	.....	250
P-6.	201	.....	.....	.....	.....	.....	250
P-7.	201	.....	.....	.....	.....	.....	250
P-8.	201	.....	.....	.....	.....	.....	250
P-9.	201	.....	.....	.....	.....	.....	250

Fig. 3: alignment by the Higgins and Sharp algorithm of eight amplicon nucleotide sequences obtained by reverse transcription polymerase chain reaction (P-1; P3-P9) and one amplicon obtained only by nested-PCR (P2) from hantavirus cardiopulmonary syndrome patients, also including the nucleotide sequence of the S segment of Araraquara virus (ARA).

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